

## Critically Appraised Topic

Syndromic testing for gastroenteritis: are conventional diagnostic methods ancient history?

# Gastroenteritis

Buikpijn, krampen en diarree (+/- koorts)

10 miljoen gevallen per jaar (België) → 28.000 hospitalisaties

Voedselvergiftigingen of dicht contact met besmette personen

**Viraal**

**Bacterieel**

**Parasitair**

Zelf-limiterend

**Richtlijnen** testen in geval van:

Aanhoudend (>7 dagen)

Acuut (<7 dagen)








Ernstig

Risicofactoren

Ouderen

Immuungecompromitteerden



BRISTOL STOOL CHART		
TYPE 1		<b>Separate hard lumps</b> VERY CONSTIPATED
TYPE 2		<b>Lumpy and sausage like</b> SLIGHTLY CONSTIPATED
TYPE 3		<b>A sausage shape with cracks in the surface</b> NORMAL
TYPE 4		<b>Like a smooth, soft sausage or snake</b> NORMAL
TYPE 5		<b>Soft blobs with clear-cut edges</b> LACKING FIBER
TYPE 6		<b>Mushy consistency with ragged edges</b> INFLAMMATION
TYPE 7		<b>Liquid consistency with no solid pieces</b> INFLAMMATION AND DIARRHEA

MEDICALNEWS TODAY

# Workflow in AZ Delta

## Allplex™ Gastrointestinal Panel Assays



*Aeromonas* spp.  
*Campylobacter* spp.  
*Salmonella* spp.  
*Shigella* spp./EIEC  
*Yersinia* spp.  
*Vibrio* spp.



*C. difficile*



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**Allplex™**  
**GI-Bacteria(II) Assay**

- EAEC<sup>[1]</sup> (*aggR*)
- EPEC<sup>[2]</sup> (*eaeA*)
- *Escherichia coli* O157 (*E. coli* O157)
- ETEC<sup>[3]</sup> (*lt/st*)
- Hypervirulent *Clostridium difficile* (CD hyper)
- STEC<sup>[4]</sup> (*stx1/2*)
- Internal Control (IC)

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**Allplex™**  
**GI-Bacteria(I) Assay**

- *Aeromonas* spp. (Aer)
- *Campylobacter* spp. (Cam)
- *Clostridium difficile* toxin B (CdB)
- *Salmonella* spp. (Sal)
- *Shigella* spp./EIEC<sup>[5]</sup> (Sh/EI)
- *Vibrio* spp. (Vib)
- *Yersinia enterocolitica* (Yer)
- Internal Control (IC)

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**Allplex™**  
**GI-Parasite Assay**

- *Blastocystis hominis* (BH)
- *Cryptosporidium* spp. (CR)
- *Cyclospora cayetanensis* (CC)
- *Dientamoeba fragilis* (DF)
- *Entamoeba histolytica* (EH)
- *Giardia lamblia* (GL)
- Internal Control (IC)

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**Allplex™**  
**GI-Virus Assay**

- Adenovirus (AdV)
- Astrovirus (AstV)
- Norovirus GI (NoV-GI)
- Norovirus GII (NoV-GII)
- Rotavirus (RotV)
- Sapovirus (SV)
- Internal Control (IC)

# Voor- en nadelen van PCR

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Lage detectiegrens

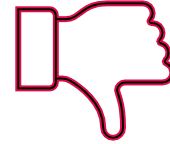
Veel **sneller** dan klassieke cultuur

**Hoge throughput** mogelijk

**Simultane detectie meerdere pathogenen**

Hoge mate van automatisatie mogelijk

Onafhankelijk van uitvoerder



**Complexe diagnostiek en interpretatie**

Geen oplossing voor antibiogram

Je vindt enkel wat in het panel zit

**Aanwezigheid  $\neq$  infectie**

Contaminatie mogelijk

**Kwalitatieve** rapportering

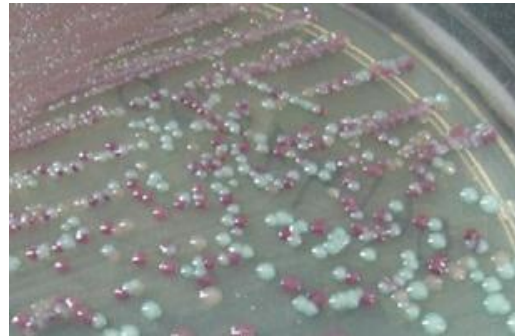
# Conventionele testwijze



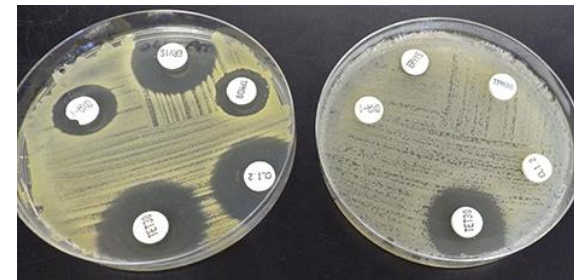
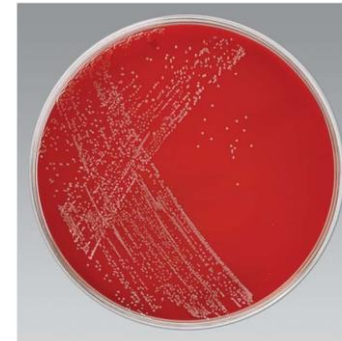
Dag 1



Dag 2



Dag 3



# Conventionele testwijze



Beperkt aantal pathogenen  
Omslachtig  
Tijdrovend  
Beperkte gevoeligheid  
Arbeidsintensief

**Klinisch beeld:** bacterieel, viraal, parasitair, niet-infectieus?

## VRAAG I

IN WELKE MATE KOMT HET AANVRAAGPROFIEL, DAT GEBASEERD OP IS  
KLINISCHE PRESENTATIE, OVEREEN MET DE RESULTATEN VAN EEN  
MULTIPLEX PCR?



# Literatuur

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## Aanvraagprofiel

In 25-65% oorzakelijke kiem **gemist** in geval van 'physician-requested testing'

## Conventionele methoden vs. PCR



High overall agreement

**Moleculaire methoden**

Aantal ↑ (tot 2x meer)

Gemengde infecties ↑

Microscopie parasieten lage gevoeligheid t.o.v. PCR

# Algemeen

1238 stalen

Conventionele methoden

267 positief (21.6%)  
281 pathogenen

Multiplex PCR

511 positief (41.3%)  
832 pathogenen

Table 1 An overview of the requested tests (conventional methods) compared to the FTD Gastrointestinal panel and both its positivity rates

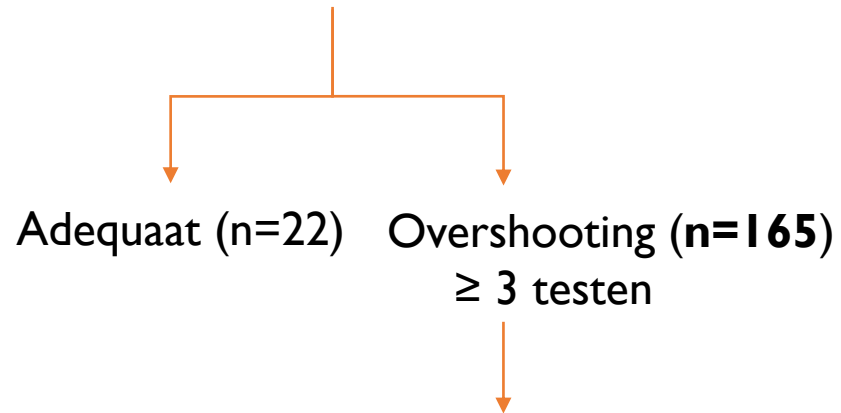
Pathogen	Conventional methods		FTD Gastrointestinal Panel	
	No. samples	No. positive	No. samples	No. positive
<b>Bacteria</b>	1222	49 (4.1%)	1238	70 (5.6%)
<b>Campylobacter</b>		29		38
<i>Campylobacter</i> spp.		1		-
<i>Campylobacter coli</i>		2		-
<i>Campylobacter jejuni</i>		26		-
<i>Salmonella</i> spp.		9		10
<i>Shigella</i> spp.		0		0
STEC		7		11
<i>Yersinia</i> spp.		5		11
<b>Viruses</b>	1009	195 (19.5%)	1238	563 (45.5%)
Adenovirus	334	105		187
Astrovirus	42*	-		58
Norovirus	378	75		161
Norovirus GI				34
Norovirus GII				127
Rotavirus	297	15		41
Sapovirus	96*	-		116
<b>Parasites</b>	434**	36 (8.3%)	1238	51 (4.1%)
<i>Cryptosporidium</i> spp.		24		36
<i>Entamoeba histolytica</i>		0		0
<i>Giardia lamblia</i>		12		15
<b><i>Clostridioides difficile</i></b>	451	35 (7.8%)	1238	148 (11.9%)

\* Amount of times adeno-, noro- and/or rotavirus was requested

\*\*404 requests for Combined Rapid Antigen Test (*Cryptosporidium* and *Giardia lamblia*) and 28 for direct microscopic examination

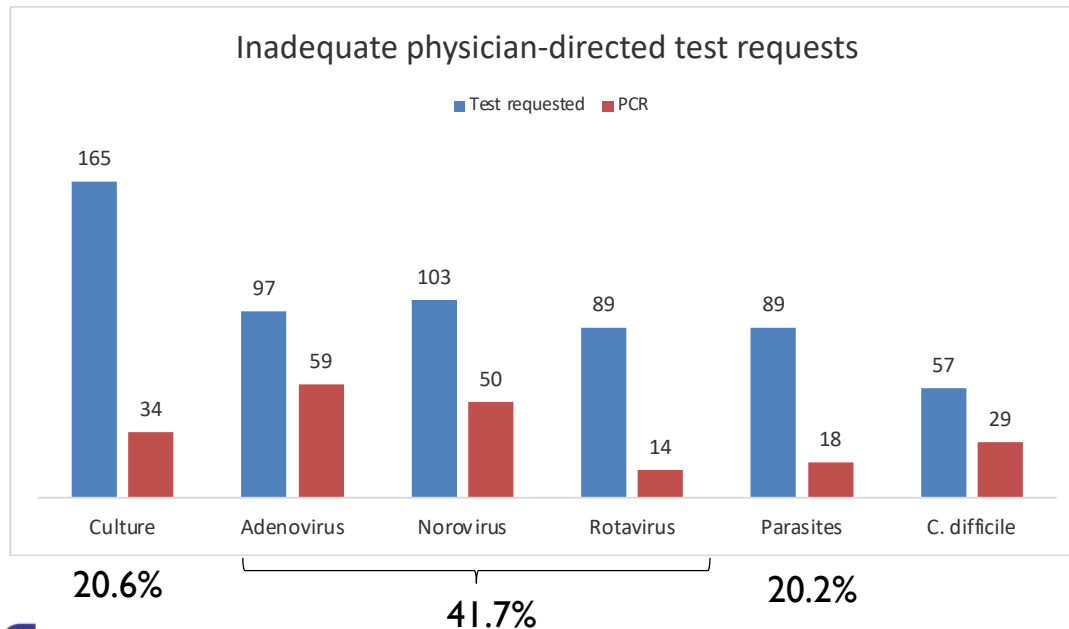
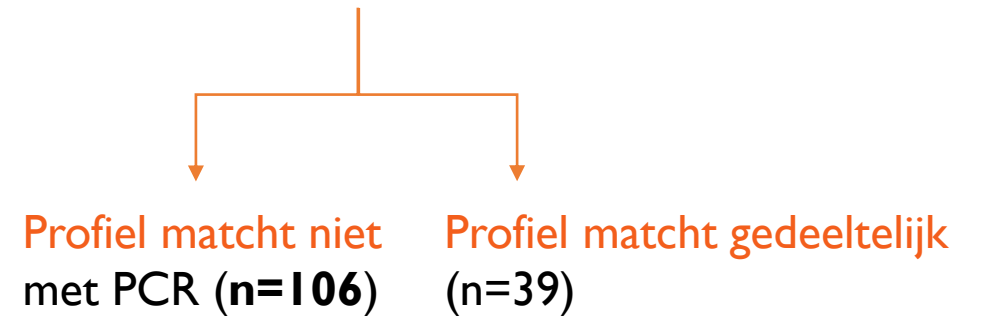
# Aanvraagprofiel

**Succesvol** (36.6%, n=187)



**Niet succesvol** (32.3%, n=165)

Exclusie 20 stalen *C. difficile* <2 jaar



# Cultuur vs. PCR

Table 3 Stool culture vs. PCR (bacteria)

	FTD positive	Stool culture positive	Stool culture negative
<b>Campylobacter spp.</b>	38	29 (76.0%)	9 (24.0%)
<b>Salmonella spp.</b>	10	9 (90.0%)	1 (10.0%)
<b>Yersinia spp.</b>	11	5 (45.5%)	6 (54.5%)
<b>STEC</b>	11	7* (63.6%)	4 (36.4%)
<b>Total</b>	70	50 (71.4%)	20 (28.6%)

\*Seven *E. coli* isolates were obtained by culture. Only 3 of these were confirmed as being a *Shiga toxin-producing E. coli* strain. Two samples were negative for Shiga toxins and for two samples no confirmation was received.

18/20 stalen Ct >30

2/20 stalen Ct 20 – 30

- Yersinia spp.
- STEC

Hoge sensitiviteit  
Geen bacteriën geïsoleerd die niet in panel zitten

# Virus antigen vs. PCR

Virale mono-infectie 178 stalen

1. **Norovirus** genotype I en II (n=60)
2. **Adenovirus** (n=44)
3. **Sapovirus** (n=33)

Table 4 Viral aetiology suspected based on clinical presentation when a *single viral pathogen* was identified by FTD

	Adenovirus (n=44)	Norovirus (n=60)	Rotavirus (n=12)
Clinical suspicion of specific viral aetiology	19 (43.2%)	19 (31.7%)	5 (41.7%)
No viral suspicion	23 (52.3%)	41 (68.3%)	6 (50.0%)

Geen vals positieve sneltesten

Table 5 Comparison between results acquired by physician-requested testing and FTD (viral)

	FTD positive	Rapid antigen test positive	Rapid antigen test negative	Rapid antigen test not requested
Adenovirus	187	107 (56.1%)	17 (9.1%)	63 (34.8%)
Norovirus GI	34	11 (32.4%)	1 (2.9%)	22 (64.7%)
Norovirus GII	127	64 (50.4%)	1 (0.8%)	62 (48.8%)
Norovirus I & II	161	75 (46.6%)	2 (1.2%)	84 (52.2%)
Rotavirus	41	15 (36.6%)	7 (17.1%)*	19 (46.3%)
<b>Total</b>	<b>389</b>	<b>197 (50.6%)</b>	<b>26 (6.7%)</b>	<b>166 (42.7%)</b>

\*Rotavirus rapid antigen test was not performed in 5 cases as the patient was older than 2 years, however 2 of them had a Ct value <20 and 3 had a Ct value >30 (FTD).

# Aanvraagprofiel

Table 2 Viral ordering profile in case of positive astrovirus or sapovirus by FTD multiplex

	Astrovirus	Sapovirus
<b>All three pathogens</b>	32/58 (55%)	68/116 (59%)
<b>Two pathogens</b>		
Adeno- and norovirus	2/58 (3.4%)	8/116 (6.9%)
Adeno- and rotavirus	3/58 (5.2%)	14/116 (12.1%)
Noro- and rotavirus	1/58 (1.7%)	0
<b>One pathogen</b>		
Adenovirus	3 (5.2%)	2 (1.7%)
Norovirus	1 (1.7%)	4 (3.4%)
Rotavirus	0	0
<b>No viral pathogen request</b>	16/58 (27.6%)	20/116 (17.2%)

# Parasieten sneltest/microscopie

Geen vals positieve sneltesten



Table 6 Comparison of results acquired by physician-requested tests for FTD (protozoa)

	FTD positive	Rapid antigen test positive	Rapid antigen test negative	Rapid antigen test not requested
<b>Cryptosporidium spp.</b>	36	24* (66.7%)	0	11 (30.6%)
<b>Giardia lamblia</b>	15	12 (80.0%)	0	3 (20.0%)
<b>Total</b>	51	36 (70.6%)		14 (27.5%)

\*One sample was a follow-up sample after therapy initiation. The rapid antigen test was not executed.

Microscopie 2/3 *Cryptosporidium* spp. gemist  
(Ct <20 en 20-30)

# Besluit

## Aanvraagprofiel

Slechts 36.6% van de aanvragen succesvol



88.2% overshooting!

## Gericht aanvragen = inadequaar

Klinische presentatie

Conventionele testen niet beschikbaar



Onderdiagnosticering

28.6% van bacteriën niet geïsoleerd

42.7% virussen niet aangevraagd

27.5% parasieten niet aangevraagd

40.0% microscopie niet gevonden

Astrovirus en sapovirus

Conventionele methoden hebben geen extra pathogenen opgeleverd in vergelijking met PCR

2 keer meer positieve stalen en pathogenen



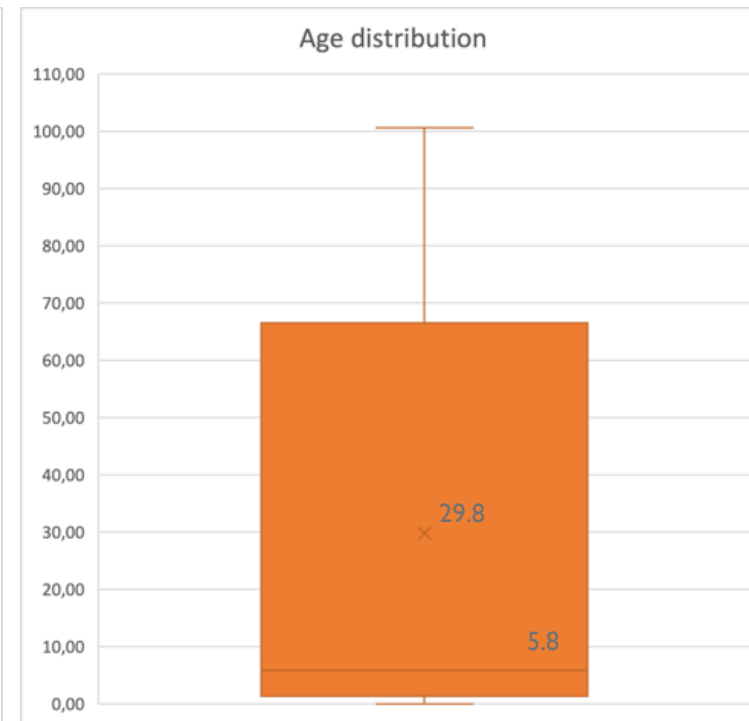
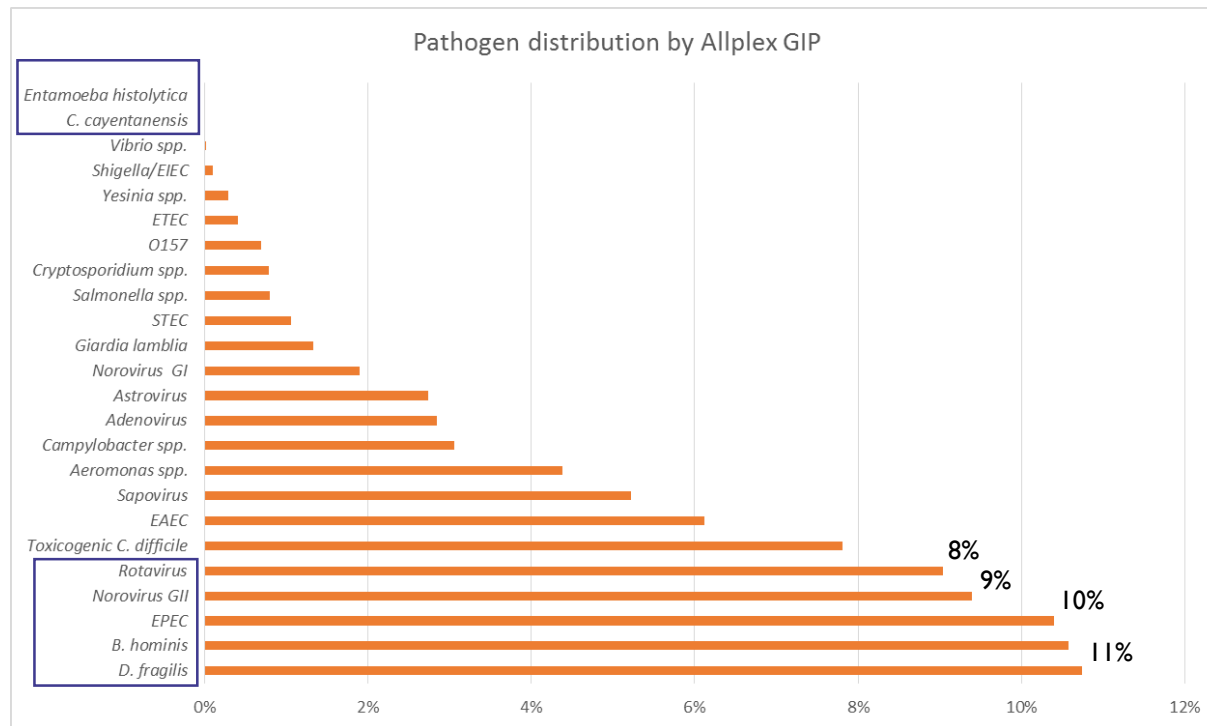


## VRAAG 2

HOE KUNNEN DE RESULTATEN GEGENEREERD DOOR SEEGENE SEMI-KWANTITATIEF GERAPPORTEERD WORDEN?

# Algemeen

8442 stalen → 4527 (53.6%) positief  
7676 pathogenen



# Algemeen

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## **Mono-infectie** 54.9%

*Blastocystis hominis* (n=448)

*Dientamoeba fragilis* (n=302)

Rotavirus (n=261)

## **Co-infectie** 45.1%

Enteropathogene *E. coli* (n=638)

Norovirus genotype II (n=571)

*Dientamoeba fragilis* (n=594)

Table 9 Total number of Seegene Allplex Gastrointestinal panel detections by age group, pathogens and co-detection

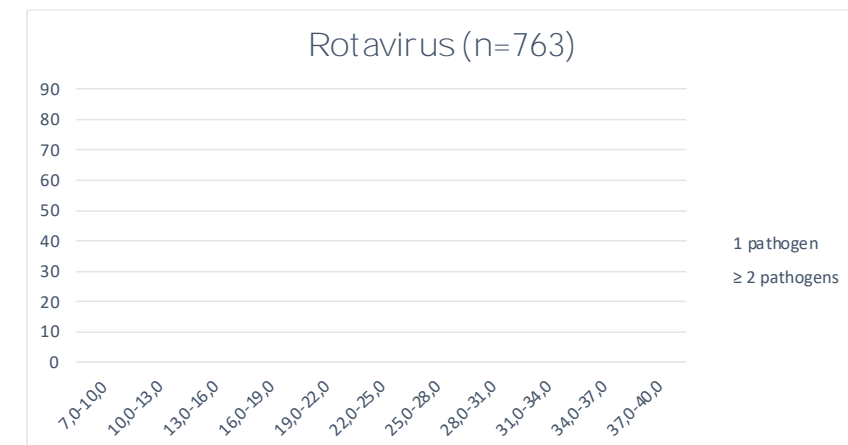
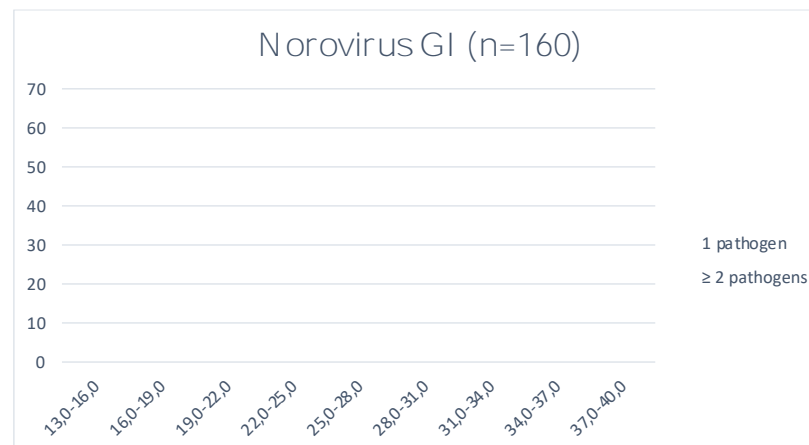
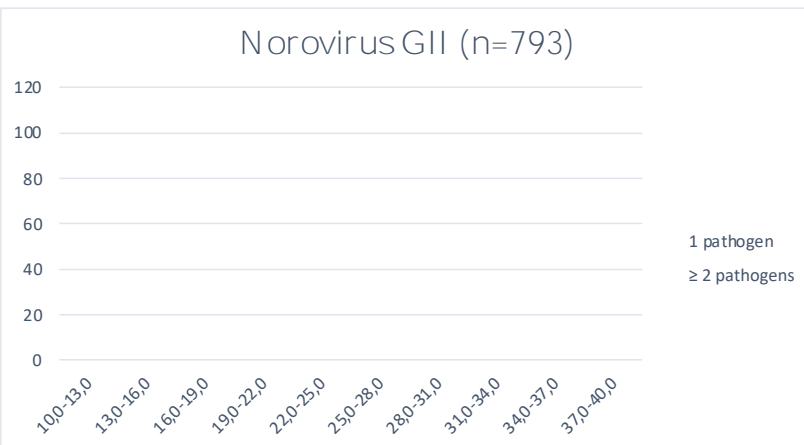
Pathogen	No (pos. ratio)	No. associated with co-infections (%)	Total no./no. associated with co-infections					
			< 1 year (n=748)	1-5 years (n=1529)	6-12 years (n=274)	13-20 years (n=109)	21-60 years (n=577)	> 60 years (n=1290)
<b>Bacteria</b>	<b>3067 (36.6%)</b>							
<i>Aeromonas</i> spp.	470 (5.6%)	331 (70.4%)	136/109	126/109	15/13	8/4	50/29	135/67
<i>Campylobacter</i> spp.	258 (3.1%)	152 (65.5%)	19/16	54/47	13/9	19/10	65/27	88/46
<i>Clostridioides difficile</i>	658* (7.8%)	431 (65.5%)	270/206	184/161	0/0	2/1	45/19	156/42
<b>E. coli</b>								
EAEC	517 (6.1%)	393 (76.0%)	111/96	280/241	7/5	5/3	38/24	76/27
EPEC	878 (10.4%)	638 (72.7%)	159/137	342/295	33/24	10/8	115/73	219/111
ETEC	35 (0.4%)	21 (60.0%)	3/3	8/8	2/2	0/0	8/5	14/4
STEC	90 (1.1%)	70 (77.8%)	8/8	16/14	5/5	5/3	16/12	40/31
O157**	59 (0.7%)	52 (88.1%)	10/8	12/12	2/2	3/3	9/7	24/22
EIEC/Shigella spp.	9 (0.1%)	5 (55.6%)	0/0	1/0	0/0	2/1	6/4	0/0
<i>Salmonella</i> spp.	68 (0.8%)	31 (45.6%)	11/5	19/10	12/5	5/2	6/5	15/4
<i>Yersinia</i> spp.	25 (0.3%)	14 (56.0%)	1/1	7/4	0/0	0/0	3/2	14/8
<i>Vibrio</i> spp.	2 (0.0%)	2 (100%)	0/0	1/1	0/0	0/0	1/1	0/0
<b>Viruses</b>	<b>2628 (31.1%)</b>							
Adenovirus	240 (2.8%)	194 (80.8%)	86/68	129/107	4/4	3/3	8/7	10/7
Astrovirus	231 (2.7%)	172 (74.5%)	65/50	137/110	4/0	1/1	13/9	11/5
Norovirus genotype I	160 (1.9%)	127 (79.4%)	26/25	86/77	5/5	3/2	9/7	31/11
Norovirus genotype II	793 (9.4%)	571 (72.0%)	220/173	403/306	20/15	10/6	69/36	71/35
Rotavirus	763 (9.0%)	497 (65.1%)	161/105	335/284	2/2	25/23	55/31	165/57
Sapovirus	441 (5.2%)	292 (66.2%)	140/100	268/179	9/6	1/0	15/5	8/3
<b>Parasites</b>	<b>1979 (23.4%)</b>							
<i>Blastocystis hominis</i>	893 (10.6%)	438 (49.0%)	4/4	101/92	89/68	48/31	185/96	466/154
<i>Cyclospora cayentanensis</i>	0 (0.0%)							
<i>Cryptosporidium</i> spp.	67 (0.8%)	44 (65.7%)	3/2	38/28	8/5	3/1	13/7	2/1
<i>Dientamoeba fragilis</i>	907 (10.7%)	594 (65.5%)	18/10	449/331	176/107	53/34	101/57	110/66
<i>Entamoeba histolytica</i>	0 (0.0%)							
<i>Giardia lamblia</i>	112 (1.3%)	77 (68.8%)	3/3	48/43	9/7	2/1	24/11	26/12

# Semi-kwantitatieve rapportering

Distributie van Ct waarden voor **elk viraal pathogeen** → semi-kwantitatieve rapportering

## Cut-off Ct waarde van 35

1. Viraal hoge Ct waarden in geval van co-infectie
2. Observatie en confirmatie van vals positieven ( $Ct > 35$ )
3. Technisch aspect



# Cut-off waarde (Ct 35)

## I. Hoge Ct waarden in geval van co-infectie

Table 10 Median Ct values of different viruses in both single and co-infections and in different patient populations

	Median Ct value single infection	Median Ct value co-infection	Median Ct value ≤ 5 years	Median Ct value >21 years
<b>Adenovirus</b>	16.89	19.71	<b>16.45</b>	<b>34.58</b>
<b>Astrovirus</b>	18.47	18.62	17.63	21.21
<b>Norovirus</b>				
Genotype I	<b>25.45</b>	<b>36.67</b>	36.04	29.47
Genotype II	<b>21.96</b>	<b>26.00</b>	24.35	28.21
<b>Rotavirus</b>	16.43	15.18	15.33	16.75
<b>Sapovirus</b>	24.46	25.63	24.90	28.01

Significant hogere Ct waarde in co-infecties

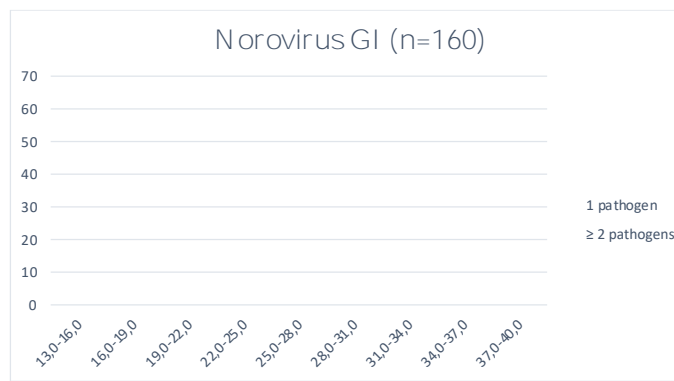
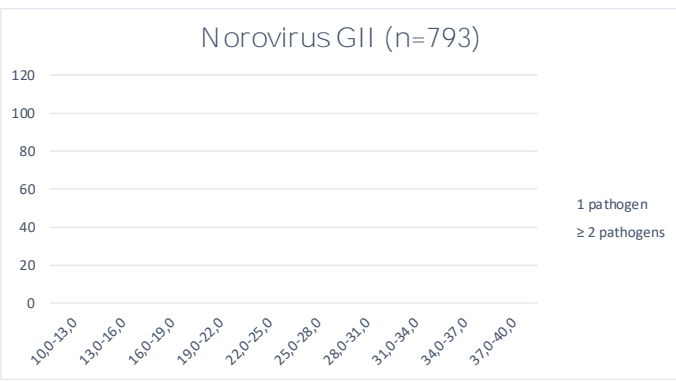
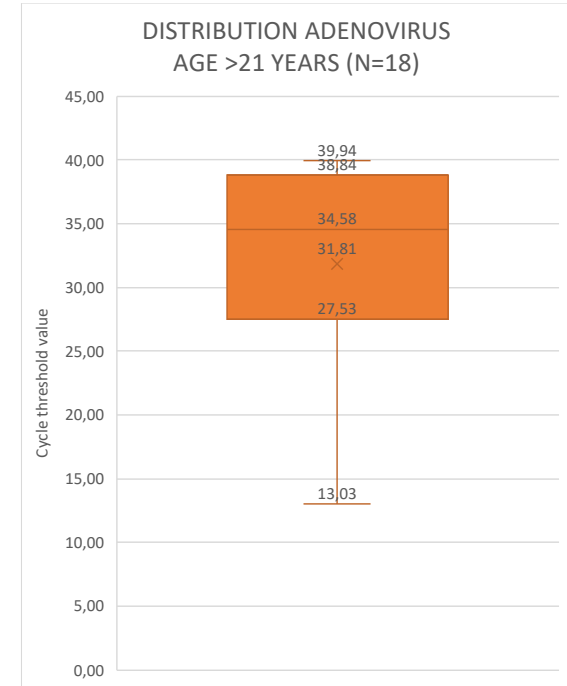
n=18

14/18 co-infectie

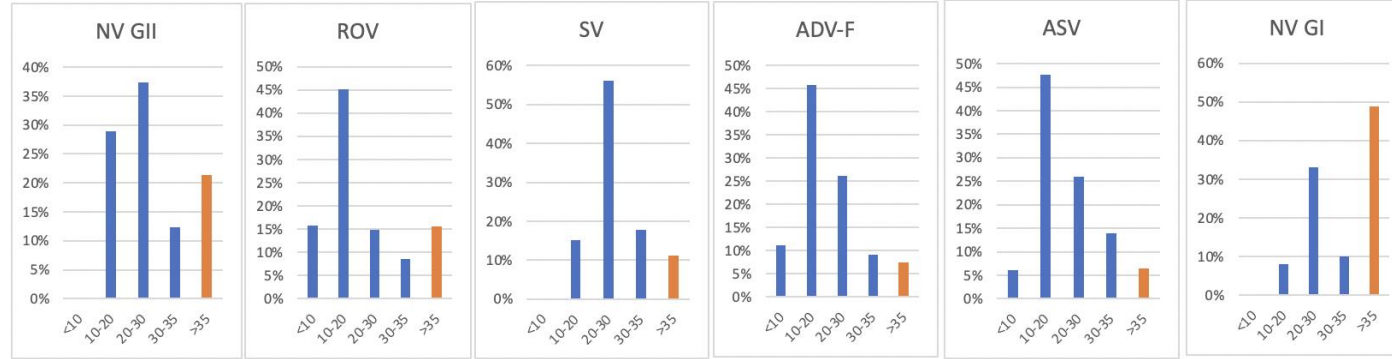
11/14 met pathogeen Ct <30

4/18 mono-infectie

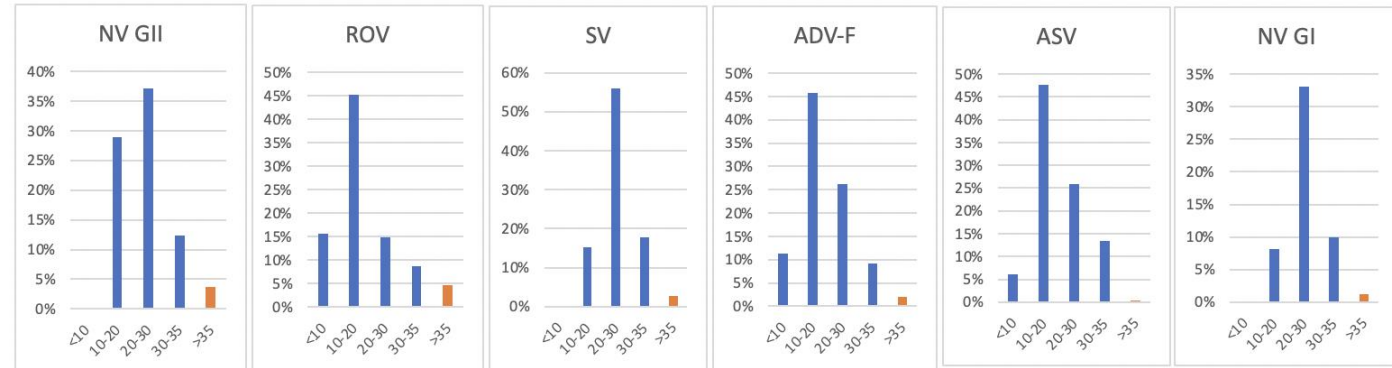
1/4 Ct <35 (13.68)

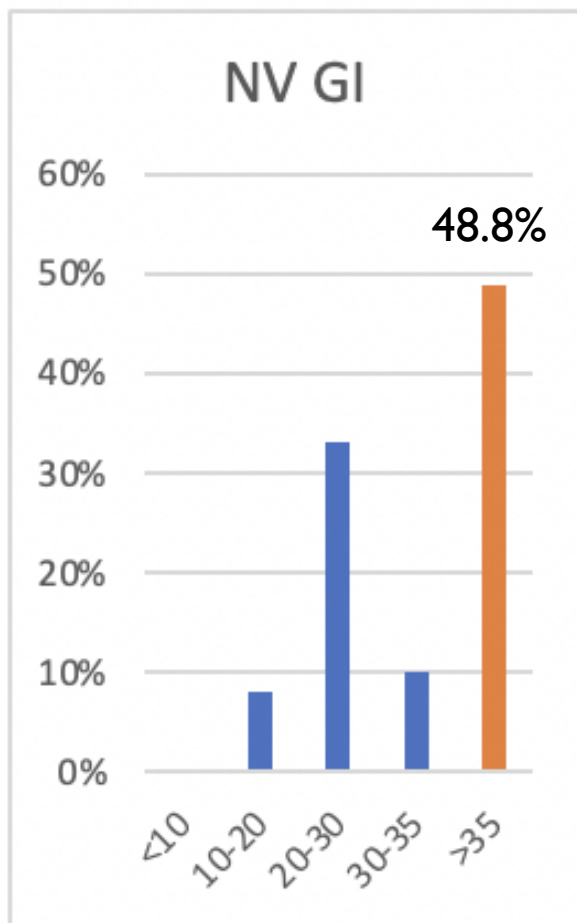


# Co-infectie met een pathogeen met Ct waarde <35



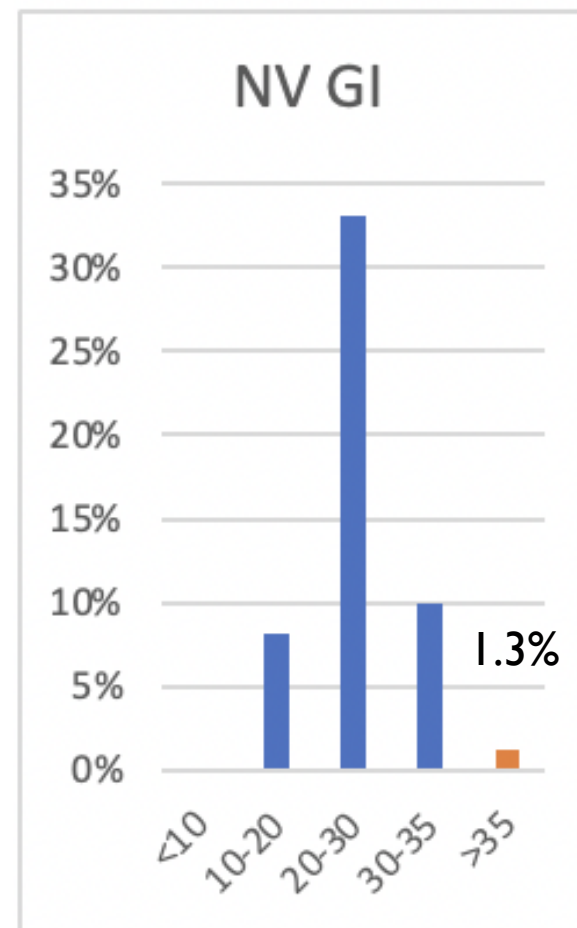
	NV GII		ROV		SV		ADV-F		ASV		NV GI	
<10	0	0.0%	120	15.7%	0	0,0%	27	11.3%	14	6.1%	0	0.0%
10-20	229	28.9%	345	45.2%	67	15.2%	110	45.8%	110	47.6%	13	8.1%
20-30	296	37.3%	113	14.8%	247	56.0%	63	26.3%	60	25.9%	53	33.1%
30-35	98	12.4%	66	8.7%	78	17.7%	22	9.2%	32	13.8%	16	10.0%
>35	170	21.4%	119	15.6%	49	11.2%	18	7.5%	15	6.5%	78	48.8%
No other pathogens	29	3.7%	36	4.72%	12	2.7%	5	2.1%	1	0.4%	2	1.3%





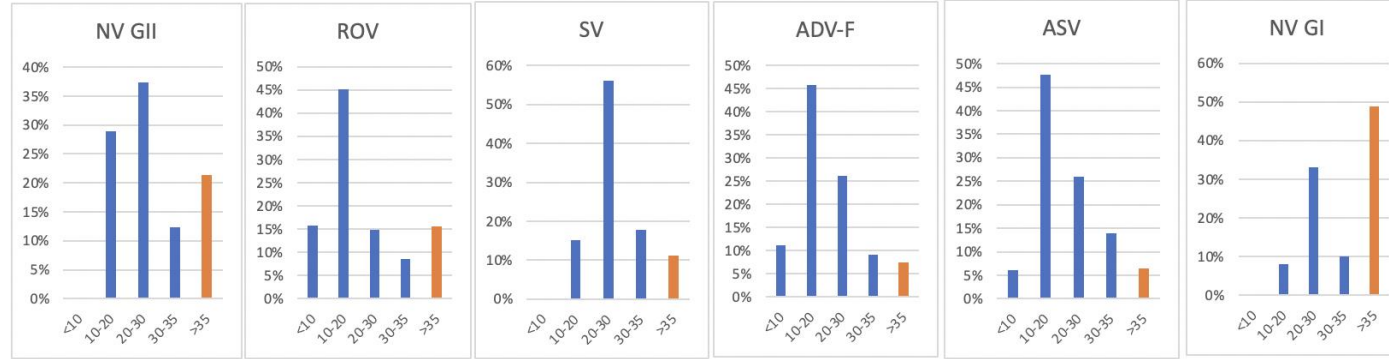
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Exclusie virale  
pathogenen met co-  
detectie met  
pathogeen Ct  
waarde <35

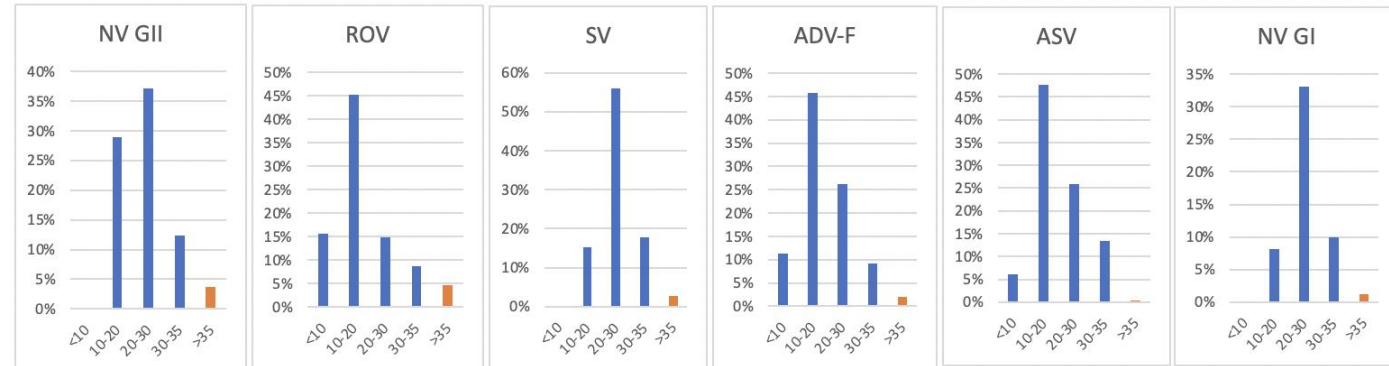




# Co-infectie met een pathogeen met Ct waarde <35



	NV GII		ROV		SV		ADV-F		ASV		NV GI	
<10	0	0.0%	120	15.7%	0	0,0%	27	11.3%	14	6.1%	0	0.0%
10-20	229	28.9%	345	45.2%	67	15.2%	110	45.8%	110	47.6%	13	8.1%
20-30	296	37.3%	113	14.8%	247	56.0%	63	26.3%	60	25.9%	53	33.1%
30-35	98	12.4%	66	8.7%	78	17.7%	22	9.2%	32	13.8%	16	10.0%
>35	170	21.4%	119	15.6%	49	11.2%	18	7.5%	15	6.5%	78	48.8%
No other pathogens	29	3.7%	36	4.72%	12	2.7%	5	2.1%	1	0.4%	2	1.3%



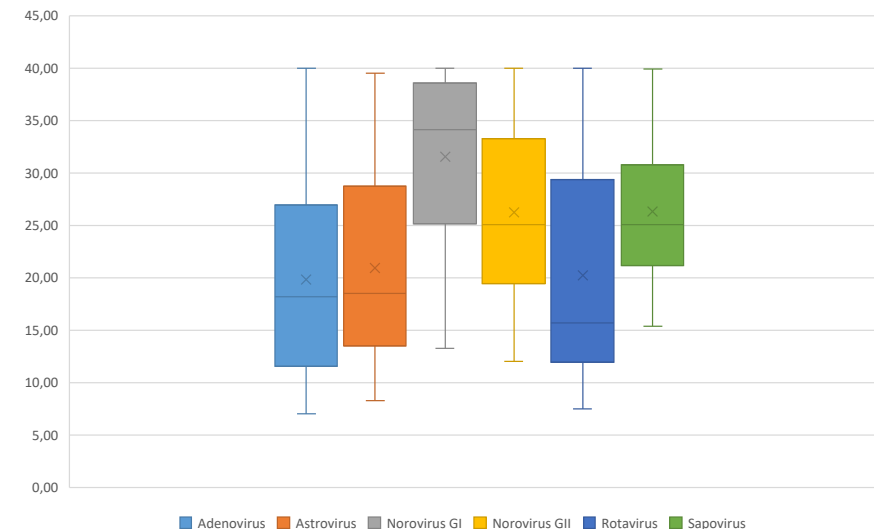
# Bepalen van cut-offs

	25th percentile	50th percentile	75th percentile
<b>Norovirus GII</b>	18.31	22.14	27.52
<b>Norovirus GI</b>	21.03	25.50	29.24
<b>Adenovirus</b>	11.12	16.34	23.84
<b>Astrovirus</b>	13.39	17.12	27.71
<b>Rotavirus</b>	10.94	14.36	21.26
<b>Sapovirus</b>	20.91	24.25	23.84



	Very strong positive	Strong positive	Positive	Weak positive
<b>Norovirus GII</b>	<18	≥18 en <22	≥22 en <28	≥28 en < 35
<b>Norovirus GI</b>	<21	≥21 en <26	≥26 en <29	≥29 en <35
<b>Adenovirus</b>	<11	≥11 en <16	≥16 en <29	≥29 en <35
<b>Astrovirus</b>	<14	≥14 en <17	≥17 en <28	≥28 en <35
<b>Rotavirus</b>	<11	≥11 en <14	≥14 en <21	≥21 en <35
<b>Sapovirus</b>	<21	≥21 en <24	≥24 en <29	≥29 en <35

OVERVIEW CT-VALUE DISTRIBUTION IN VIRUSES



# Cut-off waarde (Ct 35)

## 2. Observatie en confirmatie van vals positieve resultaten

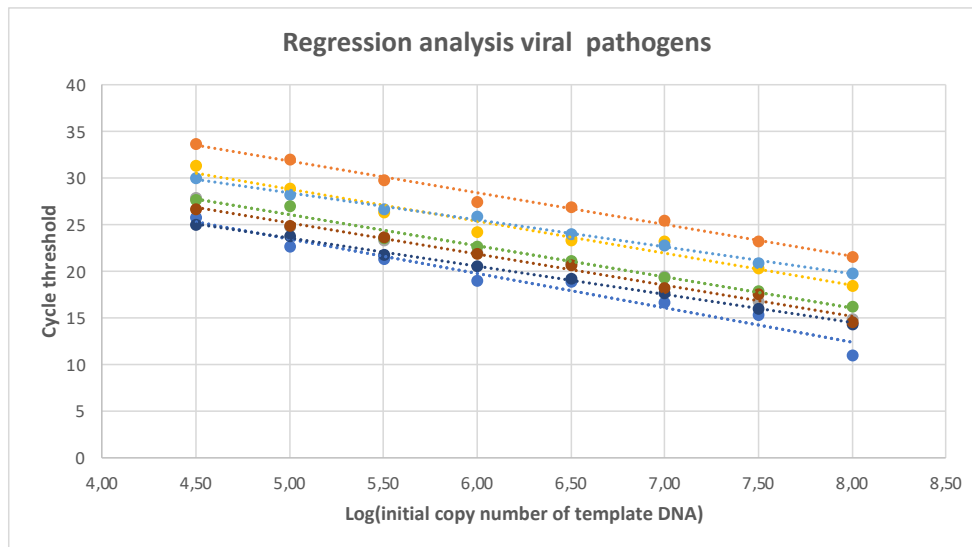
Well	Name	Type	FAM	C(t)	FAM	C(t)	HEX	C(t)	Cal Red 610	C(t)	Cal Red 610	C(t)	Quasar 670	C(t)	Quasar 670	C(t)	HEX	C(t)	Auto Interpretation
A01	2051611801	SAMPLE	ASV	C(t)	NVG2	C(t)	ADV-F	C(t)	SV	C(t)	NVG I	C(t)	ROV	C(t)		C(t)	IC	C(t)	"ADV-F,NVG I,ROV"
			-	N/A	-	N/A	+	"39,97"	-	N/A	+	"39,89"	-	"8,09"			+	"26,89"	
A06	2051611801	SAMPLE	Sh/EI	C(t)	Cam	C(t)	Yer	C(t)	Vib	C(t)	CdB	C(t)	Aer	C(t)	Sal	C(t)	IC	C(t)	"31,16"
			-	N/A	-	N/A	-	N/A	-	N/A	-	N/A	-	N/A	-	N/A	+	"31,16"	
B02	2051981301	SAMPLE	ASV	C(t)	NVG2	C(t)	ADV-F	C(t)	SV	C(t)	NVG I	C(t)	ROV	C(t)		C(t)	IC	C(t)	"NVG2,SV,NVG I,ROV"
			-	N/A	+	"39,52"	-	N/A	+	"30,98"	+	"39,51"	+	"9,51"			+	"26,75"	
B07	2051981301	SAMPLE	Sh/EI	C(t)	Cam	C(t)	Yer	C(t)	Vib	C(t)	CdB	C(t)	Aer	C(t)	Sal	C(t)	IC	C(t)	"31,47"
			-	N/A	-	N/A	-	N/A	-	N/A	-	N/A	-	N/A	-	N/A	+	"31,47"	
C01	2051794701	SAMPLE	ASV	C(t)	NVG2	C(t)	ADV-F	C(t)	SV	C(t)	NVG I	C(t)	ROV	C(t)		C(t)	IC	C(t)	"SV,NVG I,CdB"
			-	N/A	-	N/A	-	N/A	+	"20,99"	+	"37,75"	-	N/A			+	"28,27"	
C06	2051794701	SAMPLE	Sh/EI	C(t)	Cam	C(t)	Yer	C(t)	Vib	C(t)	CdB	C(t)	Aer	C(t)	Sal	C(t)	IC	C(t)	"30,59"
			-	N/A	-	N/A	-	N/A	-	N/A	+	"30,58"	-	N/A	-	N/A	+	"30,59"	
D02	2052037301	SAMPLE	ASV	C(t)	NVG2	C(t)	ADV-F	C(t)	SV	C(t)	NVG I	C(t)	ROV	C(t)		C(t)	IC	C(t)	"NVG2,ROV,CdB"
			-	N/A	+	"35,43"	-	N/A	-	N/A	-	N/A	+	"14,22"			+	"26,71"	
D07	2052037301	SAMPLE	Sh/EI	C(t)	Cam	C(t)	Yer	C(t)	Vib	C(t)	CdB	C(t)	Aer	C(t)	Sal	C(t)	IC	C(t)	"30,86"
			-	N/A	-	N/A	-	N/A	-	N/A	+	"40,90"	-	N/A	-	N/A	+	"30,86"	
G03	2052129501	SAMPLE	ASV	C(t)	NVG2	C(t)	ADV-F	C(t)	SV	C(t)	NVG I	C(t)	ROV	C(t)		C(t)	IC	C(t)	"NVG2,NVG I,ROV"
			-	N/A	+	"37,77"	-	N/A	-	N/A	+	"17,02"	+	"38,97"			+	"27,30"	
G08	2052129501	SAMPLE	Sh/EI	C(t)	Cam	C(t)	Yer	C(t)	Vib	C(t)	CdB	C(t)	Aer	C(t)	Sal	C(t)	IC	C(t)	"32,06"
			-	N/A	-	N/A	-	N/A	-	N/A	-	N/A	-	N/A	-	N/A	+	"32,06"	

Niet gedetecteerd bij herhaling met GI-TAC assay

# Virale co-infecties

8 stalen met sterk positief signaal voor één virus

↓ 0.5 log dilutie (= 1.66 Ct ↑)



Efficiëntie van 87% tot 114%  
Rico's van -3.67 tot -2.89

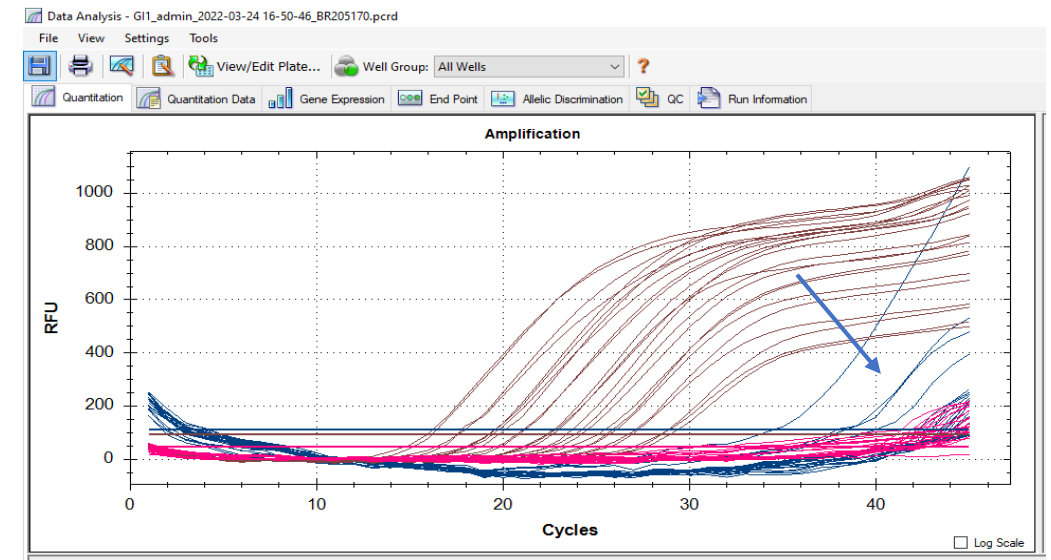
## Extra positieve signalen

Astrovirus → norovirus GII (Ct 39.65)

→ rotavirus (Ct 38.81)

Astrovirus → sapovirus (Ct 37.91)

Rotavirus (5<sup>e</sup> dilutie) → norovirus GII (Ct 38.38)



# Cut-off waarde (Ct 35)

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## 3. Technisch aspect

Minimale detectie van 100 kopie/ml staal → Ct tussen 35-40

↓  
2,5 kopieën/25µl

# Besluit

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**Cut-off** Ct waarde **35** → rapportering co-infecties ↓

Semi-kwantitatieve cut-offs zijn **verschillend per pathogeen**

**PCR** is niet foutloos!



**Geen causaal verband** aantonen

**Hoge Ct** in combinatie met lage Ct → Exclusie van oorzakelijke kiem met hoge waarschijnlijkheid

Ct is hoog of laag in een populatie met gastroenteritis

Ct als **proxy voor probabiliteit** (dus waarschijnlijk/minder waarschijnlijk als causatieve kiem) in **co-infecties**

# To do

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1. Bepalen van semi-kwantitatieve cut-off waarden voor parasieten en bacteriën
2. Dezelfde analyse doen met andere multiplex panels om ook daar semi-kwantitatief rapporteren mogelijk te maken
3. Positiviteitsratio's en Ct waarden vergelijken met een asymptomatische groep om een cut-off te kunnen stellen tussen klinisch relevant en klinisch irrelevant

# Vragen?

Don't leave your mark at the pool this summer!

It only takes one person with diarrhea to contaminate the entire pool.

Learn more at [www.cdc.gov/healthyswimming](http://www.cdc.gov/healthyswimming)

